

The Role of Mercury and Cadmium Heavy Metals in Vascular Disease, Hypertension, Coronary Heart Disease, and Myocardial Infarction

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INTRODUCTION

There is increasing concern regarding the overall health effects of exposure to various heavy metals in the environment. This is particularly true of mercury and less so with cadmium, lead, aluminum, and arsenic. The cardiovascular consequences of mercury and cadmium toxicity have not been carefully evaluated until recently. This paper will critically review the vascular consequences of mercury and cadmium toxicity in humans as it relates to hypertension, generalized atherosclerosis, coronary heart disease (CHD), myocardial infarction (MI), cerebrovascular accidents (CVA), carotid artery disease, renal dysfunction, and total mortality.

MERCURY

Types of Mercury

Mercury exists in 3 basic forms: elemental, inorganic, and organic (Table 1).^{1,5} Dental amalgams are the most common source for elemental mercury vapor, which is a stable monoatomic gas. Inorganic mercury, which is a divalent compound, is the toxic species found in human tissue after conversion from the other forms. Organic mercury in the form of methyl and ethyl mercury is primarily from fish, sea mammals, and thimerosal vaccines. Although dental amalgams historically have been the major source of human exposure, fish and sea mammals are becoming an increasingly important environmental source of potential mercury toxicity.⁶⁻⁹

TABLE 1 Mercury Types^{1,4,44}

1. Elemental	Mercury vapor (Hg ⁰) Stable monoatomic Gas	Dental amalgams
2. Inorganic	Divalent mercury (Hg ²⁺)	Toxic species in human tissue after conversion
3. Organic	Methyl mercury (CH ₃ Hg+) Ethyl mercury (CH ₃ CH ₃ Hg+)	Fish, sea mammals Thimerosal vaccines

Mercury Biotransformation and Biomethylation

Mercury from various sources, including elemental mercury from earth sources or inhaled mercury vapor, methyl and ethyl mercury, is converted by biomethylation to inorganic divalent mercury, the toxic form in human organs and tissues (Figure 1).¹⁰ Divalent mercury is soluble and stable in water and undergoes biomethylation to methyl mercury, which is found in high concentrations in certain fish and sea mammals.

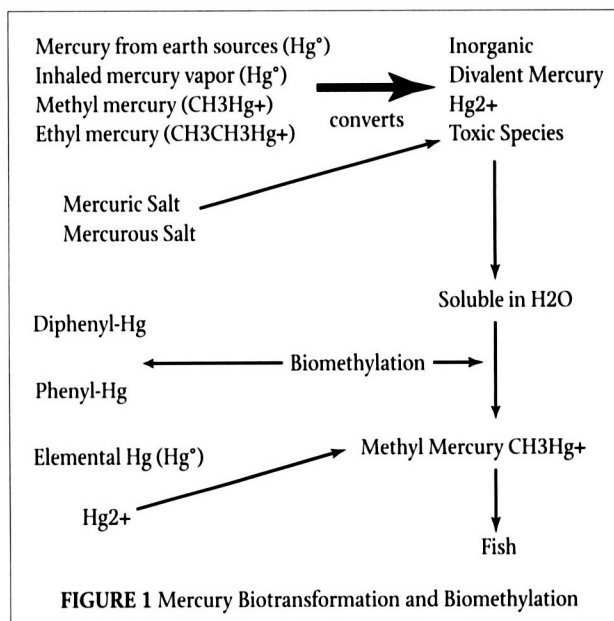


FIGURE 1 Mercury Biotransformation and Biomethylation

The Environmental Protection Agency has determined the safe daily intake of mercury to be less than 0.1 µg/kg/day (about 7 µg/day for a 154 lb person).¹¹ It is estimated that 1 dental amalgam filling releases about 3-17 µg of mercury vapor per day. The typical amalgam is composed of 50% mercury, 25% silver, 25% tin, copper, and nickel.¹²⁻¹⁴ Fish and sea mammals provide about 2-3 µg per day depending on the type and amount consumed.¹⁵⁻¹⁸ The long-lived, large predatory fish such as swordfish, tilefish, shark, and king mackerel contain about 1 µg of methyl mercury per gram. Pike, whale, bass, tuna, and trout contain about 0.1-0.5 µg of mercury per gram. Nine vaccines containing thimerosal (now off the US market) would give an estimated exposure of 62 µg of organic mercury.¹⁹⁻²² All other sources of mercury provide about 0.3 µg per day.^{23,26}

Important Facts about Mercury

Mercury is the most dangerous of all the heavy metals.²⁷ It will modify the distribution and retention of other heavy metals.²⁸⁻³⁰ Mercury has no known physiological role in human metabolism, and the human body has no mechanisms to excrete mercury actively.³¹ Mercury, thus, accumulates during life so that the average 70 kg person has a total body burden of about 13 mg of mercury.³² Mercury has a high affinity for sulfhydryl groups (-SH), various enzymes and amino acids, N-acetylcysteine (NAC), alpha lipoic acid (ALA), and glutathione (GSH), which provide about 10-50% of the plasma protein antioxidant capacity.³³⁻³⁵ Lower availability of these antioxidants reduces oxidant defense and increases oxidative stress. Selenium antagonizes some of the adverse effects of mercury by forming a seleno-mercury complex in tissue that is less toxic.³⁶⁻⁴³

Physiological Basis of Mercury Toxicity

Mercury induces mitochondrial dysfunction and oxidative stress.⁴⁴⁻⁴⁶ The primary mitochondrial dysfunction occurs at the

ubiquinone-cytochrome B region and with NADH dehydrogenase causing displacement of Fe⁺⁺ and Cu⁺ ions in the a3Cub center of cytochrome C (Figure 2). This results in depolarization and auto-oxidation of the inner mitochondrial membrane with lipid peroxidation and severe mitochondrial dysfunction. Physiologic consequences include increased hydrogen peroxide, depletion of mitochondrial glutathione by over 50%, increased lipid peroxidation markers, such as TBARS, by over 70%, oxidation of pyridine nucleotides, such as NAD(p)H, and altered calcium homeostasis.⁴⁷⁻⁴⁹ This severe mitochondrial dysfunction increases oxidative stress and reduces antioxidant defenses, creating significant health implications.

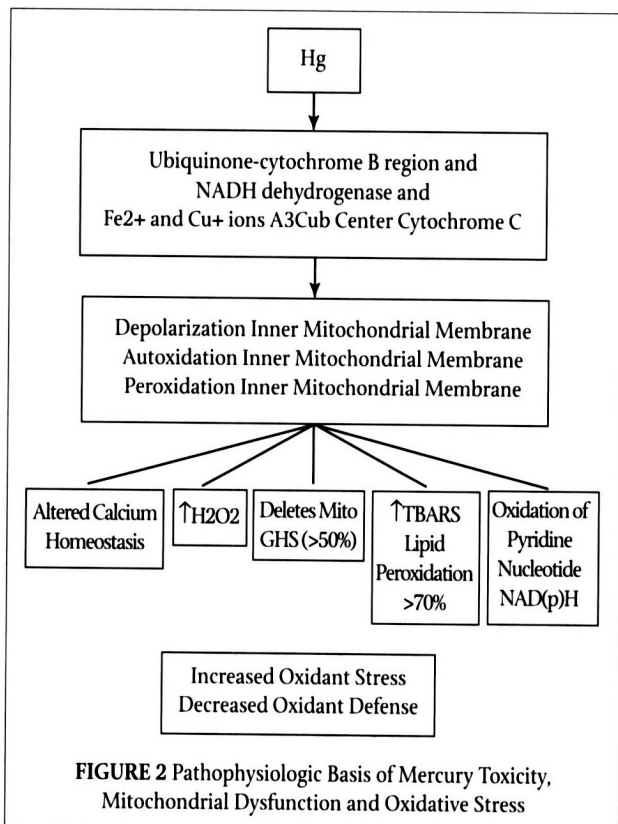


FIGURE 2 Pathophysiologic Basis of Mercury Toxicity, Mitochondrial Dysfunction and Oxidative Stress

The primary 3 sources of mercury-induced lipid peroxidation include the Fenton reaction, affinity for sulfhydryl groups, and selenium deficiency.⁵⁰ Mercury serves as a direct catalyst in Fenton-type reactions and as an indirect catalyst via iron stimulation, which increases the production of radical oxygen species and superoxide anion.⁵¹ Mercury's high affinity for sulfhydryl groups (-SH), such as glutathione, NAC, and ALA, which comprise much of the antioxidant capacity of plasma, reduces both membrane and plasma antioxidant defense. Finally, the formation of insoluble complexes of mercury with selenium reduces selenium availability, which is a necessary cofactor for glutathione peroxidase (GPx) activity to break down hydrogen peroxides and various other toxic peroxidation products. Thus, plasma and intracellular antioxidant capacity is reduced.⁵²

Vascular Biological Effects of Mercury

Numerous toxic effects of mercury have been demonstrated in vitro and in both animal and human studies (Table 2 provides details with references, and Table 3 provides a summary).

Studies find that mercury:

- Increases free radical production⁵³⁻⁶⁰

TABLE 2 Vascular Biologic Effects of Mercury

1. Increases free radical production⁵³⁻⁶⁰
2. Inactivates antioxidant defenses⁵³⁻⁵⁵
3. Binds to thiol-containing molecules^{53-55,67,68}
4. Binds to Se, forming Se-Hg complex-mercury selenide, which decreases Se available for cofactor with GPx⁵³⁻⁵⁵
5. Inactivates glutathione, catalase, and SOD⁷⁴⁻⁷⁷
6. Increases lipid peroxidation in all organs⁷⁸⁻⁸¹
7. Increases oxLDL²⁷ and oxLDL immune complexes²⁷
8. Increases platelet aggregation⁸³
9. Increases coagulation/thrombosis: increases Factor VIII PF4 and thrombin and reduces protein C^{85,86}
10. Inhibits endothelial cell formation and migration⁸⁷
11. Increases apoptosis⁸⁸
12. Reduces monocyte function and phagocytosis⁸⁸
 - Immune function is impaired
13. Increases inflammation⁸⁸

TABLE 3 Summary of Vascular Biologic Effects of Mercury

1. Oxidative stress
2. Inflammation
3. Thrombosis
4. Vascular smooth muscle (VSM) proliferation and migration
5. Endothelial dysfunction
6. Dyslipidemia (oxHDL and paraoxonase)
7. Immune dysfunction
8. Mitochondrial dysfunction

- Inactivates antioxidant defenses⁶¹⁻⁶⁴
- Binds to thiol-containing molecules⁶⁴⁻⁶⁸
- Binds to selenium, forming seleno-mercury complexes that reduce selenium availability for GPx activity⁶⁹⁻⁷³
- Inactivates glutathione, catalase, and superoxide dismutase⁷⁴⁻⁷⁷
- Increases lipid peroxidation⁷⁸⁻⁸¹
- Increases oxidation of LDL (oxLDL)
- Increases plasma oxLDL complexes⁸²

Thrombosis is potentiated by increased platelet aggregation⁸⁴ and by increases in Factor VIII, platelet factor,⁸⁴ and thrombin, with reductions in protein C.^{85,86} Endothelial cell formation and migration are reduced, which decreases vascular endothelial repair, decreases nitric oxide, and causes endothelial dysfunction.⁸⁷ Apoptosis is increased,⁸⁸ monocyte function and phagocytosis are impaired,⁸⁸ immune function is reduced,⁹⁰ and vascular inflammation is increased.⁹¹ There is an increased production and release of superoxide anion from human neutrophils and monocytes,^{92,93} depolarization of the inner mitochondrial membrane with severe mitochondrial dysfunction,⁹⁴⁻⁹⁶ and disruption of plasma membrane lipid integrity by translocation of phosphatidyl serine (PS).⁹⁷ Finally, mercury stimulates proliferation of vascular smooth muscle cells⁹⁸ and inactivates paraoxonase, an extracellular antioxidant enzyme related to HDL, CHD, and MI risk.^{99,100}

In summary, the overall vascular effects of mercury include oxidative stress, inflammation, thrombosis, VSM proliferation and migration, endothelial dysfunction, dyslipidemia, immune dysfunction, and mitochondrial dysfunction. All of these functional

abnormalities have the potential to increase the risk for hypertension and vascular disease.

Clinical Vascular Consequences of Mercury Toxicity

The clinical consequences of mercury toxicity include hypertension,¹⁰¹⁻¹⁰⁴ CHD,¹⁰⁵⁻¹⁰⁷ MI,¹⁰⁸⁻¹¹⁰ increase in carotid intimal medial thickness (IMT) and carotid obstruction,¹¹¹ CVA,¹¹² generalized atherosclerosis,¹¹³ renal dysfunction and proteinuria,¹¹⁴ and an overall increase in total mortality.¹¹⁵

Coronary Heart Disease and Myocardial Infarction

In rabbits exposed to inhaled mercury vapor, the cardiovascular and cardiac pathology included bradycardia, thrombosis in small and medium caliber arteries, focal necrosis with thickening of the endocardium of the perivalvular regions, papillary muscles and valves, and endothelial proliferation with inflammatory foci and focal edema, endothelial proliferation, inflammation, and fibrosis of the ascending aorta.¹¹⁶

In a case control study in 9 counties of 684 men with their first MI, there was a significant association of toenail mercury content, adipose tissue DHA, and first MI.¹¹⁷ There was a 15% higher toenail mercury content as assessed by neutron activation analysis (NAA) in the men with their first MI compared to the control group (95% CI: 5-25%). The risk-adjusted OR for MI was 2.16 in the highest vs the lowest quintile ($P=.006$, 95% CI: 1.09-4.29). The adipose DHA was directly proportional to the mercury toenail content ($P<.001$) and the DHA content was inversely correlated to MI with an OR of 0.59 in the highest versus the lowest quintile ($P=.02$, 95% CI: 0.30-1.19). This important study concluded that there exists a positive, monotonic increase in the risk of MI with mercury toenail content above the 0.25 $\mu\text{g/g}$ level, which was even steeper when adjusted for the DHA adipose tissue content. Mercury diminishes the cardiovascular protection of fish consumption. Another study substantiated these results—the highest quartile of DHA with the lowest quartile of mercury was associated with a 67% reduction in CHD ($P<.016$).¹¹⁸

In another large, nested case control study of 33,733 male health-care professionals between the ages of 40-75 years (Health Professionals Follow-Up Study), no association between mercury toenail content assessed by NAA and CHD was found.¹¹⁹ However, if dentists were excluded, there was a nonsignificant correlation of toenail mercury and CHD. Also, subjects with the highest tertile of mercury and the lowest serum selenium level had a significant increase in CHD.

Other human studies have shown mixed results.¹²⁰⁻¹²⁴ One study of mercury miners showed no relationship between CHD and mercury levels.¹²⁵ However, another study of European mercury miners showed a significant relationship between mercury exposure and total mortality (increase 8%), hypertension (increase 46%), CHD (increase 36%), renal disease (increase 55%), and CVA (increase 36%).¹²⁶ A Finnish study found a significant relationship between hair mercury, 24-hour urine mercury, and cardiovascular events.¹²⁷ In patients with hair mercury in the highest tertile (over 2 $\mu\text{g/g}$) and increased 24-hour urinary mercury, CHD and MI risk was increased 2-fold ($P=.005$), cardiovascular death increased by 2.9 times ($P=.014$) and circulating oxLDL and immune complexes to oxLDL increased significantly. The Gothenburg Study showed no relationship between serum mercury content and the number of amalgam fillings and CHD or MI.¹²⁸

Carotid Atherosclerosis

High hair mercury content correlates with increased carotid IMT and carotid atherosclerosis.¹²⁹ A study of 1,014 men between the ages of 42-60 years found an increase in mean carotid IMT over 4

years ($P=.0007$). Each increase of 1 μg in hair content equaled an 8 μmol increase in carotid IMT, a 7.3% increase over the mean. There was a 0.042 mm/4-year difference in the highest quintile versus the lowest quintile, which correlated to a 32% greater increase ($P<.05$). In addition, mercury hair content was proportional to blood pressure, fibrinogen levels, waist-hip ratio, and low HDL cholesterol; all were significant at $P=.0002$.

Hypertension

The association of mercury toxicity and hypertension in humans is convincing.¹³⁰⁻¹³³ Mercury miners were found to have significant increases in systolic blood pressure ($P<.01$) that correlated with lipid peroxidation and overall oxidative stress ($P<.01$).¹³⁴ European mercury miners had a 46% greater incidence of hypertension vs aged-matched controls. Other studies have shown significant correlations with hair mercury content, hypertension, and carotid IMT.¹³⁵

In acute and probably chronic mercury intoxication, mercury binds to the sulfhydryl group S-adenosyl methionine (S-AdoMet) and inactivates this enzyme, which is a necessary cofactor for catecholamine-O-methyl transferase (COMT), the enzyme needed to convert norepinephrine, epinephrine, and dopamine by methoxylation.¹³⁶ This results in a clinical syndrome that resembles a pheochromocytoma crisis with malignant hypertension in acute mercury intoxication and significant increases in urinary catecholamines in chronic mercury toxicity. This can be a very helpful clinical clue to mercury-induced hypertension. Mercury also induces renal dysfunction and proteinuria, which contribute to sodium retention and hypertension.¹³⁷⁻¹⁴⁰ Studies have shown an increase in renal insufficiency in mercury miners of 55%.¹⁴¹ Mercury concentrates in the renal tubules and glomerulus and results in proteinuria, fibrosis, and chronic renal insufficiency and dysfunction.^{142,143}

CADMIUM

The role of cadmium in cardiovascular disease and hypertension is less convincing than that of mercury due to methodological flaws and study design in most of the published human studies.¹⁴⁴⁻¹⁵¹ Cadmium exposure is uncommon in most of the population unless there is oral consumption of polluted water or chronic inhalation exposure from cigarettes.¹⁵² Twenty cigarettes will release about 30 μg of cadmium, of which 2-4 μg is actually inhaled.¹⁵³ The oral absorption of cadmium in tap water is 13-19% or about 2-4 μg per day.¹⁵⁴ Absorption of cadmium is increased in the presence of low dietary calcium, iron, and protein.¹⁵⁵ Cadmium concentrates in all organs, but mostly in kidney, liver, and pancreas, and has a half-life of over 30 years in renal tissue.¹⁵⁶

Serum and urinary cadmium reflect recent exposure, but not total body burden.¹⁵⁷ Cadmium binds to metallothionein,¹⁵⁸ substitutes for zinc and copper in metalloenzymes,¹⁵⁹ and has a high affinity for sulfhydryl groups, similar to mercury.¹⁶⁰

Animal studies show that cadmium toxicity causes aortic and coronary atherosclerosis, reduces cardiac output, alters the cardiac conduction system, reduces ATP, increases cholesterol and free fatty acids, increases blood pressure, and induces renal tubular dysfunction, proteinuria, and chronic renal insufficiency.¹⁶¹⁻¹⁶⁵ These effects are mitigated by calcium administration.

Human studies attempting to show a relationship between cadmium and cardiovascular disease or hypertension are subject to many methodological errors, so that accurate conclusions are difficult to draw.¹⁶⁶⁻¹⁶⁸ In human autopsy studies, there is a poor correlation between renal cadmium content and hypertension.¹⁶⁹ In those studies where hypertension and cadmium coexist, the mechanisms include

increases in urinary catecholamines, renal toxicity with proteinuria, sodium retention, increased intracellular calcium, and alteration in Na⁺/K⁺ ATPase.¹⁷⁰ Cadmium concentrates in the renal cortex and tubules and there reduces the expression of renal cortical CYP4A11.¹⁷¹ CYP4A11 is involved in the hydroxylation of PUFA and affects sodium balance through 20 HETE. The combination of increased renal tubular sodium reabsorption, direct renal toxicity, and proteinuria increases the risk of hypertension.¹⁷²

Cadmium also increases metallothionein in renal tubular cells and other tissues, which alters intracellular zinc.¹⁷³ This reduces zinc-dependent ligand binding to DNA and reduces PPAR expression, and may increase free fatty acids, lipids, glucose, and blood pressure. It is possible that some degree of insulin resistance occurs, which contributes to many of the associated metabolic disturbances noted above.

It is quite likely that chronic high exposure to cadmium in smokers, those drinking polluted water, and those with the CYP4A11 genetic alteration could have cadmium-induced hypertension and cardiovascular disease, but additional human studies are required to confirm this association.

SUMMARY

1. Mercury, cadmium, and other heavy metals have a high affinity for sulfhydryl (-SH) groups, inactivating numerous enzymatic reactions, amino acids, and sulfur-containing antioxidants (NAC, ALA, GSH), with subsequent decreased oxidant defense and increased oxidative stress. Both bind to metallothionein and substitute for zinc, copper, and other trace metals reducing the effectiveness of metalloenzymes.

2. Mercury induces mitochondrial dysfunction with reduction in ATP, depletion of glutathione, and increased lipid peroxidation; increased oxidative stress is common.

3. Selenium antagonizes mercury toxicity.

4. The overall vascular effects of mercury include oxidative stress, inflammation, thrombosis, vascular smooth muscle dysfunction, endothelial dysfunction, dyslipidemia, immune dysfunction, and mitochondrial dysfunction.

5. The clinical consequences of mercury toxicity include hypertension, CHD, MI, increased carotid IMT and obstruction, CVA, generalized atherosclerosis, and renal dysfunction with proteinuria. Pathological, biochemical, and functional medicine correlations are significant and logical.

6. Mercury diminishes the protective effect of fish and omega-3 fatty acids.

7. Mercury, cadmium, and other heavy metals inactivate COMT, which increases serum and urinary epinephrine, norepinephrine, and dopamine. This effect will increase blood pressure and may be a clinical clue to heavy metal toxicity.

8. Cadmium concentrates in the kidney, particularly inducing proteinuria and renal dysfunction; it is associated with hypertension, but less so with CHD. Renal cadmium reduces CYP4A11 and PPARs, which may be related to hypertension, sodium retention, glucose intolerance, dyslipidemia, and zinc deficiency. Dietary calcium may mitigate some of the toxicity of cadmium.

9. Heavy metal toxicity, especially mercury and cadmium, should be evaluated in any patient with hypertension, CHD, or other vascular disease. Specific testing for acute and chronic toxicity and total body burden using hair, toenail, urine, serum, etc. with baseline and provoked evaluation should be done.

References

1. Keating MH, Mahaffey KR, Schoemy R, et al. *Mercury study report to Congress, Vol. I, Executive summary. EPA-452/R-97-003*. Washington, DC: Environmental Protection Agency;1997.

2. Committee on the Toxicological Effects of Methylmercury, Board on Environmental Studies and Toxicology, Commission on Life Sciences. *Toxicological effects of methylmercury*. Washington, DC: National Research Council; 2000.
3. Magos L. Physiology and toxicology of mercury. *Met Ions Biol Syst*. 1997;34:321-370.
4. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
5. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
6. Keating MH, Mahaffey KR, Schoemy R, et al. *Mercury study report to Congress, Vol. I, Executive summary. EPA-452/R-97-003*. Washington, DC: Environmental Protection Agency;1997.
7. Committee on the Toxicological Effects of Methylmercury, Board on Environmental Studies and Toxicology, Commission on Life Sciences. *Toxicological effects of methylmercury*. Washington, DC: National Research Council; 2000.
8. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
9. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
10. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
11. Ibid.
12. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
13. Ahlqvist M, Bengtsson C, Lapidus L, Gergdahl IA, Schutz A. Serum mercury concentration in relation to survival, symptoms, and diseases: results from the prospective population study of women in Gothenburg, Sweden. *Acta Odontol Scand*. 1999;57(3):168-174.
14. Bergdahl IA, Schutz A, Ahlqvist M, et al. Methylmercury and inorganic mercury in serum—correlation to fish consumption and dental amalgam in a cohort of women born in 1922. *Environ Res*. 1998;77(1):20-24.
15. Keating MH, Mahaffey KR, Schoemy R, et al. *Mercury study report to Congress, Vol. I, Executive summary. EPA-452/R-97-003*. Washington, DC: Environmental Protection Agency;1997.
16. Committee on the Toxicological Effects of Methylmercury, Board on Environmental Studies and Toxicology, Commission on Life Sciences. *Toxicological effects of methylmercury*. Washington, DC: National Research Council; 2000.
17. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
18. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
19. Keating MH, Mahaffey KR, Schoemy R, et al. *Mercury study report to Congress, Vol. I, Executive summary. EPA-452/R-97-003*. Washington, DC: Environmental Protection Agency;1997.
20. Committee on the Toxicological Effects of Methylmercury, Board on Environmental Studies and Toxicology, Commission on Life Sciences. *Toxicological effects of methylmercury*. Washington, DC: National Research Council; 2000.
21. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
22. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
23. Keating MH, Mahaffey KR, Schoemy R, et al. *Mercury study report to Congress, Vol. I, Executive summary. EPA-452/R-97-003*. Washington, DC: Environmental Protection Agency;1997.
24. Committee on the Toxicological Effects of Methylmercury, Board on Environmental Studies and Toxicology, Commission on Life Sciences. *Toxicological effects of methylmercury*. Washington, DC: National Research Council; 2000.
25. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
26. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
27. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
28. Yoshizawa K, Rimm EB, Morris JS, et al. Mercury and the risk of coronary heart disease in men. *N Engl J Med*. 2002;347(22):1755-1760.
29. Chmielnicka J, Bem EM, Kaszubski P. Organ and subcellular distribution of cadmium in rats exposed to cadmium, mercury, and selenium. *Environ Res*. 1983;31(2):266-272.
30. Komsta-Szumaska E, Chmielnicka J. Effects of zinc, cadmium or copper on mercury distribution in rat tissues. *Toxicol Lett*. 1983;17(3-4):349-354.
31. International Programme on Chemical Safety (IPCS). *Methylmercury. Environmental Health Criteria 101*. Geneva: World Health Organization; 1990.
32. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
33. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
34. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.
35. International Programme on Chemical Safety (IPCS). *Methylmercury. Environmental Health Criteria 101*. Geneva: World Health Organization. 1990.

36. Yoshizawa K, Rimm EB, Morris JS, et al. Mercury and the risk of coronary heart disease in men. *N Engl J Med*. 2002;347(22):1755-1760.
37. Parizek J, Ostadalova I. The protective effect of small amounts of selenite in sublethal intoxication. *Experientia*. 1967;23(2):142-143.
38. Ganther HE, Goudie C, Sunde ML, Kopecky MJ, Wagner P. Selenium: relation to decreased toxicity of methylmercury added to diets containing tuna. *Science*. 1972;175(26):1122-1124.
39. Ganther HE, Sunde ML. Effect of tuna fish and selenium on the toxicity of methylmercury: a progress report. *J Food Sci*. 1974;39:1-5.
40. Stoewsand GS, Bache CA, Lisk DJ. Dietary selenium protection of methylmercury intoxication of Japanese quail. *Bull Environ Contam Toxicol*. 1974;11(2):152-156.
41. Sumino K, Yamamoto R, Kitamura SA. A Role of selenium against methylmercury toxicity. *Nature*. 1977;268(5615):73-74.
42. Singhal RK, Anderson ME, Meister A. Glutathione, a first line of defense against cadmium toxicity. *FASEB J*. 1987;1(3):220-223.
43. Seppanen K, Kantola M, Laatikainen R, et al. Effect of supplementation with organic selenium on mercury status as measured by mercury in pubic hair. *J Trace Elem Med Biol*. 2000;14(2):84-87.
44. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
45. Shenker BJ, Guo TL, Shapiro IM. Low-level methylmercury exposure causes human T-cells to undergo apoptosis: evidence of mitochondrial dysfunction. *Environ Res*. 1998;77(2):149-159.
46. Peraza MA, Ayala-Fierro F, Barber DS, Casarez E, Rael LT. Effects of micronutrients on metal toxicity. *Environ Health Perspect*. 1998;106(Suppl 1):203-216.
47. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
48. Shenker BJ, Guo TL, Shapiro IM. Low-level methylmercury exposure causes human T-cells to undergo apoptosis: evidence of mitochondrial dysfunction. *Environ Res*. 1998;77(2):149-159.
49. Peraza MA, Ayala-Fierro F, Barber DS, Casarez E, Rael LT. Effects of micronutrients on metal toxicity. *Environ Health Perspect*. 1998;106(Suppl 1):203-216.
50. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
51. Ibid.
52. Ibid.
53. Magos L. Physiology and toxicology of mercury. *Met Ions Biol Syst*. 1997;34:321-370.
54. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
55. Clarkson TW. The toxicology of mercury. *Crit Rev Clin Lab Sci*. 1997;34(4):369-403.
56. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
57. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
58. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidative capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
59. Park ST, Lim KT, Chung YT, Kim SU. Methylmercury-induced neurotoxicity in cerebral neuron culture is blocked by antioxidants and NMDA receptor antagonists. *Neurotoxicology*. 1996;17(1):37-45.
60. Miller DM, Lund BO, Woods JS. Reactivity of Hg(II) with superoxide: evidence for the catalytic dismutation of superoxide by Hg(II). *J Biochem Toxicol*. 1991;6(4):293-298.
61. Magos L. Physiology and toxicology of mercury. *Met Ions Biol Syst*. 1997;34:321-370.
62. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
63. Clarkson TW. The toxicology of mercury. *Crit Rev Clin Lab Sci*. 1997;34(4):369-403.
64. Magos L. Physiology and toxicology of mercury. *Met Ions Biol Syst*. 1997;34:321-370.
65. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
66. Clarkson TW. The toxicology of mercury. *Crit Rev Clin Lab Sci*. 1997;34(4):369-403.
67. Naganuma A, Koyama Y, Imura N. Behavior of methylmercury in mammalian erythrocytes. *Toxicol Appl Pharmacol*. 1980;54(3):405-410.
68. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
69. Magos L. Physiology and toxicology of mercury. *Met Ions Biol Syst*. 1997;34:321-370.
70. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
71. Clarkson TW. The toxicology of mercury. *Crit Rev Clin Lab Sci*. 1997;34(4):369-403.
72. Cuvin-Aralar ML, Furness RW. Mercury and selenium interaction: a review. *Ecotoxicol Environ Saf*. 1991;21(3):348-364.
73. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidative capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
74. Naganuma A, Koyama Y, Imura N. Behavior of methylmercury in mammalian erythrocytes. *Toxicol Appl Pharmacol*. 1980;54(3):405-410.
75. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
76. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
77. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidative capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
78. Rungby J, Ernst E. Experimentally induced lipid peroxidation after exposure to chromium, mercury or silver: interactions with carbon tetrachloride. *Pharmacol Toxicol*. 1992;70(3):205-207.
79. Park ST, Lim KT, Chung YT, Kim SU. Methylmercury-induced neurotoxicity in cerebral neuron culture is blocked by antioxidants and NMDA receptor antagonists. *Neurotoxicology*. 1996;17(1):37-45.
80. Huang YL, Cheng SL, Lin TH. Lipid peroxidation in rats administered with mercuric chloride. *Biol Trace Elem Res*. 1996;52(2):193-206.
81. Lin TH, Huang YL, Huang SF. Lipid peroxidation of rats administered with methylmercuric chloride. *Biol Trace Elem Res*. 1996;54(1):33-41.
82. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
83. Kostka B. Kinetic evaluation of ADP-induced platelet aggregation potentiation by methylmercuric chloride. *J Trace Elem Exp Med*. 1991;4:1-9.
84. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
85. Kostka B. Kinetic evaluation of ADP-induced platelet aggregation potentiation by methylmercuric chloride. *J Trace Elem Exp Med*. 1991;4:1-9.
86. Wierzbicki R, Prazanowski M, Michalska M, Krajewska U, Mieliwicki WP. Disorders in blood coagulation in humans occupationally exposed to mercuric vapors. *J Trace Elem Exp Med*. 2002;15(1):21-29.
87. Kishimoto T, Oguri T, Abe M, Kajitani H, Tada M. Inhibitory effect of methylmercury on migration and tube formation by cultured human vascular endothelial cells. *Arch Toxicol*. 1995;69(6):357-361.
88. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
89. Ibid.
90. Ibid.
91. Ibid.
92. Jansson G, Harms-Ringdahl M. Stimulating effects of mercuric- and silver ions on the superoxide anion production in human polymorphonuclear leukocytes. *Free Radic Res Commun*. 1993;18(2):87-98.
93. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
94. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
95. Shenker BJ, Guo TL, Shapiro IM. Low-level methylmercury exposure causes human T-cells to undergo apoptosis: evidence of mitochondrial dysfunction. *Environ Res*. 1998;77(2):149-159.
96. Peraza MA, Ayala-Fierro F, Barber DS, Casarez E, Rael LT. Effects of micronutrients on metal toxicity. *Environ Health Perspect*. 1998;106(Suppl 1):203-216.
97. Insug O, Datar S, Koch CJ, Shapiro IM, Shenker BJ. Mercuric compounds inhibit human monocyte function by inducing apoptosis: evidence for formation of reactive oxygen species, development of mitochondrial membrane permeability transition and loss of reductive reserve. *Toxicology*. 1997;124(30):211-224.
98. Lu KP, Zhao SH, Wang DS. The stimulatory effect of heavy metal cations on proliferation of aortic smooth muscle cells. *Sci China B*. 1990;33(3):303-310.
99. Gonzalez MC, Gil F, Hernandez AF, Villanueva E, Pla A. Inhibition of paraoxonase activity in human liver microsomes by exposure to EDTA, metals and mercurials. *Chem Biol Interact*. 1997;105(3):169-179.
100. Salonen JT, Malin R, Tuomainen TP, Nyyssonen K, Lakka TA, Lehtimaki T. Polymorphism in high density lipoprotein paraoxonase gene and risk of acute myocardial infarction in men: prospective nested case-control study. *BMJ*. 1999;319(7208):487-488.
101. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
102. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.

103. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidant capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
104. Torres AD, Rai AN, Hardiek ML. Mercury intoxication and arterial hypertension: report of two patients and review of the literature. *Pediatrics*. 2000;105(3):E34.
105. Barregard L, Sallsten G, Jarvholm B. Mortality and cancer incidence in chloralkali workers exposed to inorganic mercury. *Br J Ind Med*. 1990;47(2):99-104.
106. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
107. Ganther HE, Goudie C, Sunde ML, Kopecky MJ, Wagner P. Selenium: relation to decreased toxicity of methylmercury added to diets containing tuna. *Science*. 1972;175(26):1122-1124.
108. Barregard L, Sallsten G, Jarvholm B. Mortality and cancer incidence in chloralkali workers exposed to inorganic mercury. *Br J Ind Med*. 1990;47(2):99-104.
109. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
110. Singhal RK, Anderson ME, Meister A. Glutathione, a first line of defense against cadmium toxicity. *FASEB J*. 1987;1(3):220-223.
111. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.
112. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001; 58:461-466.
113. Clarkson TW, Magos L, Myers GJ. The toxicology of mercury—current exposures and clinical manifestations. *N Engl J Med*. 2003;349(18):1731-1737.
114. Ibid.
115. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
116. Wojciechowski J, Kowalski W. Cardiac and aortic lesions in chronic experimental poisoning with mercury vapors. *Pol Med Sci Hist Bull*. 1975;15(2):255-260.
117. Guallar E, Sanz-Gallardo MI, van't Veer P, et al. Mercury, fish oils, and the risk of myocardial infarction. *N Engl J Med*. 2002;347(22):1747-1754.
118. Rissanen T, Voutilainen S, Nyyssonen K, Lakka TA, Salonen JT. Fish oil-derived fatty acids, docosahexaenoic acid and docosapentaenoic acid, and the risk of acute coronary events: the Kuopio Ischaemic Heart Disease Risk Factor Study. *Circulation*. 2000;102:2677-2679.
119. Yoshizawa K, Rimm EB, Morris JS, et al. Mercury and the risk of coronary heart disease in men. *N Engl J Med*. 2002;347(22):1755-1760.
120. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
121. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
122. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidant capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
123. Torres AD, Rai AN, Hardiek ML. Mercury intoxication and arterial hypertension: report of two patients and review of the literature. *Pediatrics*. 2000;105(3):E34.
124. Ahlqwist M, Bengtsson C, Lapidus L, Gergdahl IA, Schutz A. Serum mercury concentration in relation to survival, symptoms, and diseases: results from the prospective population study of women in Gothenburg, Sweden. *Acta Odontol Scand*. 1999;57(3):168-174.
125. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidant capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
126. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
127. Salonen JT, Seppanen K, Nyyssonen K, et al. Intake of mercury from fish, lipid peroxidation, and the risk of myocardial infarction and coronary, cardiovascular, and any death in eastern Finnish men. *Circulation*. 1995;91(3):645-655.
128. Ahlqwist M, Bengtsson C, Lapidus L, Gergdahl IA, Schutz A. Serum mercury concentration in relation to survival, symptoms, and diseases: results from the prospective population study of women in Gothenburg, Sweden. *Acta Odontol Scand*. 1999;57(3):168-174.
129. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.
130. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
131. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.
132. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidant capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
133. Torres AD, Rai AN, Hardiek ML. Mercury intoxication and arterial hypertension: report of two patients and review of the literature. *Pediatrics*. 2000;105(3):E34.
134. Kobal AB, Horvat M, Prezelj M, et al. The impact of long-term past exposure to elemental mercury on antioxidant capacity and lipid peroxidation in mercury miners. *J Trace Elem Med Biol*. 2004;17(4):261-274.
135. Salonen JT, Seppanen K, Lakka TA, Salonen R, Kaplan GA. Mercury accumulation and accelerated progression of carotid atherosclerosis: a population-based prospective 4-year follow-up study in men in eastern Finland. *Atherosclerosis*. 2000;148(2):265-273.
136. Torres AD, Rai AN, Hardiek ML. Mercury intoxication and arterial hypertension: report of two patients and review of the literature. *Pediatrics*. 2000;105(3):E34.
137. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
138. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
139. Kobal AB, Flisar Z, Miklavcic V, Dizdarevic T, Sesek-Briski A. Renal function in miners intermittently exposed to elemental mercury vapour. *Arh Hig Rada Toksikol* 2000;51(4):369-380.
140. Torres AD, Rai AN, Hardiek ML. Mercury intoxication and arterial hypertension: report of two patients and review of the literature. *Pediatrics*. 2000;105(3):E34.
141. Boffetta P, Sallsten G, Garcia-Gomez M, et al. Mortality from cardiovascular diseases and exposure to inorganic mercury. *Occup Environ Med*. 2001;58(7):461-466.
142. Lund BO, Miller DM, Woods JS. Studies on Hg(II)-induced H₂O₂ formation and oxidative stress in vivo and in vitro in rat kidney mitochondria. *Biochem Pharmacol*. 1993;45(10):2017-2024.
143. Kobal AB, Flisar Z, Miklavcic V, Dizdarevic T, Sesek-Briski A. Renal function in miners intermittently exposed to elemental mercury vapour. *Arh Hig Rada Toksikol* 2000;51(4):369-380.
144. Revis NW, Zinsmeister AR, Bull R. Atherosclerosis and hypertension induction by lead and cadmium ions: an effect prevented by calcium ion. *Proc Natl Acad Sci U S A*. 1981;78(10):6494-6498.
145. Hallenbeck WH. Human health effects of exposure to cadmium. *Experientia*. 1984;40(2):136-142.
146. Spieker C, Zidek W, Zumkley H. Cadmium and hypertension. *Nephron*. 1987;47(Suppl 1):34-36.
147. Kopp SJ, Perry HM Jr, Perry EF, Erlanger M. Cardiac physiologic and tissue metabolic changes following chronic low-level cadmium and cadmium plus lead ingestion in the rat. *Toxicol Appl Pharmacol*. 1983;69(1):149-160.
148. Kristensen TS. Cardiovascular diseases and the work environment. A critical review of the epidemiologic literature on chemical factors. *Scand J Work Environ Health*. 1989;15:245-264.
149. Baker JR, Satarug S, Edwards RJ, Moore MR, Williams DJ, Reilly PE. Potential for early involvement of CYP isoforms in aspects of human cadmium toxicity. *Toxicol Lett*. 2003;137(1-2):85-93.
150. Brzoska MM, Moniuszko-Jakoniuk J. The influence of calcium content in diet on cumulation and toxicity of cadmium in the organism. *Arch Toxicol*. 1998;72:63-73.
151. Subramanyam G, Bhaskar M, Govindappa S. The role of cadmium in induction of atherosclerosis in rabbits. *Indian Heart J*. 1992;44(3):177-180.
152. Hallenbeck WH. Human health effects of exposure to cadmium. *Experientia*. 1984;40(2):136-142.
153. Ibid.
154. Ibid.
155. Ibid.
156. Ibid.
157. Ibid.
158. Ibid.
159. Brzoska MM, Moniuszko-Jakoniuk J. The influence of calcium content in diet on cumulation and toxicity of cadmium in the organism. *Arch Toxicol*. 1998;72:63-73.
160. Ibid.
161. Revis NW, Zinsmeister AR, Bull R. Atherosclerosis and hypertension induction by lead and cadmium ions: an effect prevented by calcium ion. *Proc Natl Acad Sci U S A*. 1981;78(10):6494-6498.
162. Hallenbeck WH. Human health effects of exposure to cadmium. *Experientia*. 1984;40(2):136-142.
163. Kopp SJ, Perry HM Jr, Perry EF, Erlanger M. Cardiac physiologic and tissue metabolic changes following chronic low-level cadmium and cadmium plus lead ingestion in the rat. *Toxicol Appl Pharmacol*. 1983;69(1):149-160.
164. Brzoska MM, Moniuszko-Jakoniuk J. The influence of calcium content in diet on cumulation and toxicity of cadmium in the organism. *Arch Toxicol*. 1998;72:63-73.
165. Subramanyam G, Bhaskar M, Govindappa S. The role of cadmium in induction of atherosclerosis in rabbits. *Indian Heart J*. 1992;44(3):177-180.
166. Hallenbeck WH. Human health effects of exposure to cadmium. *Experientia*. 1984;40(2):136-142.
167. Spieker C, Zidek W, Zumkley H. Cadmium and hypertension. *Nephron*. 1987;47(Suppl 1):34-36.
168. Kristensen TS. Cardiovascular diseases and the work environment. A critical review of the epidemiologic literature on chemical factors. *Scand J Work Environ Health*. 1989;15:245-264.
169. Hallenbeck WH. Human health effects of exposure to cadmium. *Experientia*. 1984;40(2):136-142.
170. Kristensen TS. Cardiovascular diseases and the work environment. A critical review of the epidemiologic literature on chemical factors. *Scand J Work Environ Health*. 1989;15:245-264.
171. Baker JR, Satarug S, Edwards RJ, Moore MR, Williams DJ, Reilly PE. Potential for early involvement of CYP isoforms in aspects of human cadmium toxicity. *Toxicol Lett*. 2003;137(1-2):85-93.
172. Ibid.
173. Ibid.